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## Review Article

# Anticancer Activity of Green Tea Polyphenols in Prostate Gland

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Numerous evidences from prevention studies in humans, support the existence of an association between green tea polyphenols consumption and a reduced cancer risk. Prostate cancer is one of the most frequently diagnosed male neoplasia in the Western countries, which is in agreement with this gland being particularly vulnerable to oxidative stress processes, often associated with tumorigenesis. Tea polyphenols have been extensively studied in cell culture and animal models where they inhibited tumor onset and progression. Prostate cancer appears a suitable target for primary prevention care, since it grows slowly, before symptoms arise, thus offering a relatively long time period for therapeutic interventions. It is, in fact, usually diagnosed in men 50-year-old or older, when even a modest delay in progression of the disease could significantly improve the patients quality of life. Although epidemiological studies have not yet yielded conclusive results on the chemopreventive and anticancer effect of tea polyphenols, there is an increasing trend to employ these substances as conservative management for patients diagnosed with less advanced prostate cancer. Here, we intend to review the most recent observations relating tea polyphenols to human prostate cancer risk, in an attempt to outline better their potential employment for preventing prostate cancer.

## 1. Introduction

In the past decade, prostate cancer (PCa) has been one of the most frequently diagnosed male neoplasias in the Western countries, and despite recent important progress, it continues to represent a major cause of cancer-related mortality. The reasons of this high incidence are unknown. Racial and ethnic differences in PCa incidence and mortality are well known, with African-American men being at the greatest risk for diagnosis, followed by Caucasian and Hispanic men. Asian-Americans seem to be at the lowest risk for PCa [1].

Generally, PCa appears to be sporadically inherited (less than 10%). These observations highlight the hypothesis that interactions between multiple genetic and ambient factors are significative determinant in PCa development.

Diet is believed one of the most probable and determinant environmental risk factors. The hypothesis results were strengthened by ecological studies showing that the PCa incidence rapidly increases in Asian immigrants that have assimilated Western diet and way of living, and in Asian men that, although living in their original countries, are contaminated by Western lifestyle, tending to substitute soy, tea, fish,

fruits, and vegetables consumption with red meat and fatty food [2]. The molecular mechanisms, through which racial, genetic, environmental conditions affect PCa development, are still a matter of discussion.

Numerous experimental evidences suggest that both dietary and lifestyle factors act by promoting chronic inflammation and/or oxidative stress leading to DNA damage, epigenetic modifications, or other alterations associated with cancer initiation. Altogether, the experimental data so far produced suggest that antioxidant and anti-inflammatory agents may play a promising role for PCa prevention [3]. In fact, (1) the proliferative inflammatory atrophy (PIA) has been proposed as a precursor to prostatic intraepithelial neoplasia (PIN) that merges with high-grade PIN (HGPIN) in about 34% of cancerous lesions [4]. Chronic inflammation may damage epithelial cells and lead to proliferative lesions, likely PIN lesions, and prostatic carcinomas precursors [5]; (2) several evidences have suggested that oxidative stress, following from the imbalance of reactive oxygen species (ROS) production and cellular antioxidant defences, is one of the most critical aging-associated factor on prostate carcinogenesis. Cumulative ROS effect possibly results in lipids, proteins, and DNA damage [6]. Prostate gland is known to be particularly vulnerable to oxidative stress, probably because of inflammation and hormonal deregulation processes and epigenetic modifications, frequently occurring in the organ.

It is worth to underline that PCa is a suitable target for primary chemopreventive interventions, since it is a unique malignancy that generally grows very slowly, before symptoms arise. As a consequence, it offers a relatively long time period for therapeutic interventions and, because of its long latency, it is typically diagnosed in 50-year-old men or older, when even a modest delay in the disease progression could significantly improve the patient quality of life.

Considering that most of the known chemotherapeutic treatments against PCa carry side effects risk, there is an increasing trend to employ conservative management for patients diagnosed with less advanced PCa, that may not require treatment. These types of tumors, in fact, are relatively indolent, almost never relapse after local therapy, and probably require a simple watchful waiting.

In order to obtain new additional opportunities improving nontoxic chemopreventive strategies, dietary substances consumption, especially tea polyphenols, can represent an important clinical challenge.

Tea, the most popular worldwide consumed beverage after water, obtained from the dried leaves of the plant *Camellia sinensis*, has been studied extensively for its effects on cancer prevention. The major polyphenols in green tea, generally known as catechins, (-)-epigallocatechin-3-gallate (EGCG), (-)-epigallocatechin (EGC), (-)-epicatechin-3-gallate (ECG), and (-)-epicatechin (EC), display meta-5,7-dihydroxyl groups on the A ring [7] and di- or trihydroxyl groups on the B ring, that represent the principal site of antioxidant reactions [8]. EGCG and ECG, harboring D ring (gallate), present maximal antioxidant activity (Figure 1). The above characteristics allow tea polyphenols to react with ROS (superoxide radical, singlet oxygen, hydroxyl/peroxyl radical, peroxynitrite) [9] and prevent, as strong metal ion

chelators, ROS output following several compounds auto-oxidation. Due to the ability of acting as good donors for hydrogen bonding, an accurate prediction about tea polyphenols solubility and permeability remains, at the moment, an elusive target [10].

Tea polyphenols have been shown to inhibit tumorigenesis and tumor progression, at different organ sites, in different animal models for human cancer. Many evidences highlighted that these compounds affect enzyme activities and signal transduction pathways, resulting in cell proliferation suppression, apoptosis enhancement, as well as angiogenesis and cell invasion inhibition, finally inhibiting the development of the disease.

To date, an association between green tea polyphenols consumption and reduced cancer risk is also supported by human cancer prevention studies, although epidemiological evidences have not yet yielded conclusive results on their chemopreventive and anticancer effect against PCa, possibly owing to different confounding factors [11].

## 2. Mechanisms of GTCs Action in PCa Cell Lines

Several mechanisms involved in GTCs inhibition of cancer formation/progression are recently reviewed by numerous Authors [12–15]. Certainly, GTCs, through their antioxidant activity, are able to quench ROS and chelate transition metals, produced during all the carcinogenesis stage. However, it has been reported that also GTCs can be a source of ROS generation, inducing oxidative stress and consequently activating apoptotic pathways [16].

GTCs, and especially EGCG, are capable of modulating a plethora of cell signalling pathways crucial for cancer cells transformation and survival, including, but not limited to, the mitogen-activated protein kinase (MAP-kinase), the nuclear factor-kappa B (NFκB), and the insulin-like growth factor (IGF)/IGF-1 receptor pathways.

With regard to the prostate-specific processes GTCs are able to affect androgen receptor (AR) downregulation and prostate-specific antigen (PSA) expression [14, 17].

Here below, we report the most probable GTCs mechanisms of action in some PCa cell lines.

### 2.1. Inhibition of Cell Proliferation and Cell Cycle Arrest.

GTCs exhibit ant-proliferative effects versus both androgen-sensitive and androgen-insensitive human PCa cells. The effect is mediated by cell cycle deregulation and cell death induction [18].

We showed that GTCs action is cancer specific, since GTCs is capable of inducing growth arrest both in SV-40 immortalized prostate epithelial cells (PNT1a) and in tumorigenic androgen-independent PCa cells (PC3), while normal human prostate epithelial cells were not significantly affected, even when EGCG was administered at higher doses [19]. The IC<sub>50</sub> of EGCG ranges from about 40 to about 200 μM, depending on the cell line type (LNCap < PNT1a < DU145 < PC3), as well as the length of the experiment, ranging from 24 to 72 hours [18, 20]. Our results were confirmed by other authors in normal fibroblasts [20, 21].

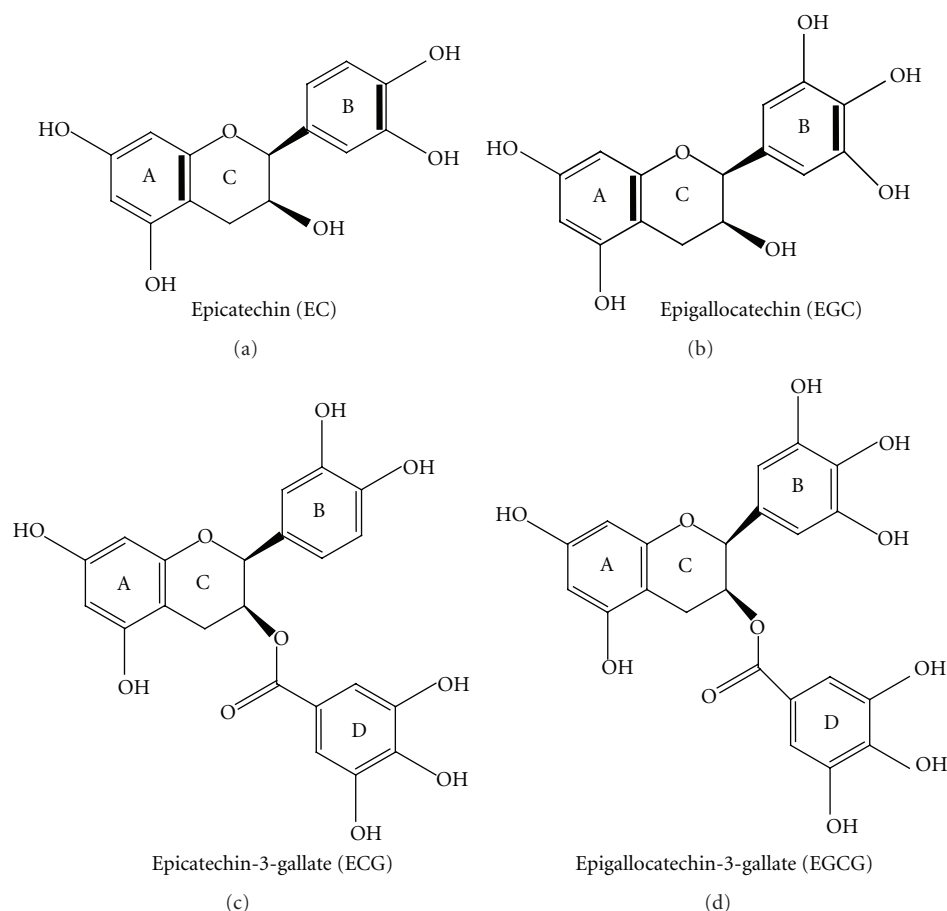


FIGURE 1: Chemical structure of the green tea catechins obtained from the dried leaves of the plant *Camellia sinensis*.

Both in LnCap and DU145 cells, EGCG treatment resulted in the cell cycle arrest at G<sub>0</sub>/G<sub>1</sub> phase (dose dependent) and the cyclin kinase inhibitor WAF1/p21 and apoptosis induction, irrespective of the AR presence or p53 cell status. Cells harbouring an active p53 protein respond to lower EGCG doses than cells with a mutated p53 [18].

It was shown that EGCG treatment of LNCaP and DU145 cells causes an induction of G<sub>1</sub>-phase cyclin kinase inhibitors, that, in turn, inhibit the cyclin-cyclin-dependent kinase (CDK) complexes operative in the G<sub>0</sub>/G<sub>1</sub> phase, thereby causing a cell cycle arrest, a possible irreversible process ultimately leading to apoptotic cell death [22].

**2.2. Apoptosis.** Hastak et al. [23] reported that EGCG induces apoptosis in LNCaP cells mainly through modulation of two related pathways: (i) p53 stabilization by specific phosphorylation and downregulation of murine double minute 2 protein (MDM2), mediated by the tumor suppressor p14ARF; (ii) negative regulation of NFκB activity, which results in reduction of the proapoptotic Bcl2 family protein expression. EGCG-induced p53 stabilization causes up-regulation of its downstream targets WAF1/p21 and Bax; consequently, EGCG produces a change in the Bax/Bcl2 ratio, shifting the balance between pro-/antiapoptotic proteins in

favor of apoptosis. This first event triggers the caspases cascade activation, followed by poly-ADP-Ribose polymerase (PARP) cleavage and chromatin fragmentation. Inactivation of p53, by using Small Interfering RNA, renders LnCap cells more resistant to EGCG-mediated apoptosis. On the other hand, stable transfection of PC3 cells (that are endowed with a mutated and inactivated p53 protein) with a cDNA encoding wild-type p53, allows to by-pass their resistance to EGCG-mediated apoptosis [24]. Ablation of p21 or Bax confers a growth advantage to the cells through inhibition of the mitochondrial pathway of caspase-dependent apoptosis.

Tumour necrosis factor-related apoptosis-inducing ligand/Apo2L ligand (TRAIL/Apo2L) is believed a promising candidate for cancer therapy, even if emergence of drug resistance limiting its potential use occurs.

Siddiqui et al. [25] reported that EGCG treatment sensitizes TRAIL-resistant LNCaP cells to TRAIL-mediated apoptosis through modulation of apoptotic pathways. TRAIL/Apo2L, when combined with EGCG, exhibited enhanced cells apoptotic activity, characterized by three major molecular events: (i) increase of PARP cleavage; (ii) proantiapoptotic Bcl2 family proteins modulation, favoring apoptosis; (iii) synergistic inhibition of apoptosis inhibitors and concomitant increase in caspase activation.

**2.3. Anti- or Pro-Oxidant Effects and Activation of Phase II Detoxifying Enzymes.** GTCs were shown to suppress cell growth and induce apoptosis in DU145, through increasing ROS formation and mitochondrial depolarization. Although the molecular mechanisms are still not clear, GTCs-induced apoptosis is not related to the members of Bcl2 family, as EGCG did not alter Bcl2, BclX(L) and BAD expression, in this cell line [26].

Nuclear factor-E2-related factor 2 (Nrf2) is a transcription factor that plays a pivotal role in the antioxidant response and oxidative stress, through activation of phase II detoxifying or antioxidant enzymes [27, 28]. EGCG, as well as other electrophile natural compounds, are able to activate a core antioxidant responsive element (EpRe), present in the promoter region of many genes involved in the cellular response to oxidative stress [28, 29]. A widely accepted model for induction of EpRe-mediated antioxidant gene expression, plausible also for EGCG, involves phosphorylation of Nrf2, leading to enhanced Nrf2 accumulation and subsequent EpRe binding [28].

Activator Protein-1 (AP-1) is a redox-sensitive transcription factor that transduces modifications of the cellular redox status, modulating the expression of genes, including pro-survival genes, in responses to oxidative and electrophilic stresses.

Nair et al. [30] demonstrated that EGCG, when administered to PC3 cells together with sulforaphane (SFN) is able to reduce AP-1 induction. The authors also confirmed, by “*in silico*” analyses, the presence of conserved transcription factor binding site in the Nrf2 and AP-1 promoter region, suggesting that gene expression changes induced by SFN and EGCG, could be mediated via concerted modulation of the Nrf2 and AP-1 pathways [30].

**2.4. Modulation of the NF $\kappa$ B Signalling and Inhibition of Inflammation Pathways.** NF $\kappa$ B is a redox sensitive transcription factor, often overexpressed in tumor and cancer cell lines, that has been suggested to regulate a variety of cellular functions, including inflammation, immune response, growth, and cell death. NF $\kappa$ B resides in the cytoplasm, bound to its inhibitor I $\kappa$ B; once the inhibitor is released, NF $\kappa$ B phosphorylation occurs, followed by its translocation to the nucleus.

EGCG has been shown to decrease the DNA binding activity of NF $\kappa$ B, and reduce the expression of the p65 subunit of NF $\kappa$ B in LNCaP cells, stimulated by tumour necrosis factor  $\alpha$ . NF $\kappa$ B over-expression is an important target in PCa due to the regulation of various downstream targets that include the cyclooxygenase-2 proteins (COX-2) [23].

In the LNCaP (androgen-dependent) and PC3 (androgen independent) prostate cancer cells, EGCG was shown to inhibit mitogen stimulated COX-2 expression through a mechanism probably involving the regulation of transcription factors, like NF $\kappa$ B, and not the direct binding to the enzyme [31]. Moreover, LNCaP and PC3 cells treatment with a combination of EGCG and COX-2 inhibitors resulted in: (i) enhanced cell growth inhibition; (ii) caspase-dependent induction of apoptosis and PARP cleavage; (iii) inhibition

of peroxisome proliferator-activated receptor gamma; (iv) NF $\kappa$ B inhibition, when compared with the effects of the two singularly employed agents, suggesting that they play a synergistic role [32].

Matrix metalloproteinases (MMPs), generally involved in extracellular matrix degradation, result overexpressed in PCa and play an important role in tumor progression and invasion. NF $\kappa$ B is the key transcription factor involved in the regulation of MMPs genes.

In DU145 cells Vayalil and Katiyar [33] demonstrated that EGCG inhibits MMP-2 and MMP-9 (inactive and active form expression) through a dose-dependent phosphorylation inhibition of the extracellular signal-regulated kinase (ERK1/2) and p38 pathways. Inhibition of the activation of transcription factors c-jun and NF $\kappa$ B also occurs.

Siddiqui et al. [25] observed that EGCG, administered in combination with TRAIL, can inhibit LNCaP cells invasion and migration potential. The authors found the effect is mediated through inhibition of the expression of the following factors: vascular endothelial growth factor (VEGF), urokinase plasminogen activator (uPA), and angiopoietin 1 and 2. A significant inhibition in both MMP-2 and -9 protein expression and activity occurs, in the presence of up-regulation of the tissue inhibitor of metalloproteinases 1.

**2.5. Insulin-Like Growth Factor (IGF) Axis, Mitogen-Activated Protein (MAP) Kinases Pathway, Phosphoinositide-3 Kinase (PI3K/AKT) Pathway.** It has been suggested that the IGF axis plays a relevant role in PCa onset and development. Binding of IGF1 to its cognate receptor activates the intracellular tyrosine kinase domain, that produces phosphorylation of many protein substrates, including members of the MAP kinase cascade and PI3K/AKT. MAP kinases and the PI3K/AKT pathway are both involved in the complex modulation of signalling pathways, which regulates cellular processes like proliferation, survival/death, and motility, usually altered in carcinogenesis.

It has been reported that EGCG inhibits IGF-1 receptor activity with an IC<sub>50</sub> of 14  $\mu$ M [34]. Treatment of DU145 and LNCaP cells with subapoptotic EGCG doses reduces IGF-induced growth [35].

Siddiqui et al. [36] found that EGCG is able to: (i) decrease PI3K and phospho-Akt levels and (ii) increase ERK1/2 level in both DU145 and LNCaP cells. Treatment of PC3 cells with EGCG results in activation of the ERK1/2 pathway, that is, not dependent by mitogen-extracellular signal-regulated kinase (MEK), the immediate upstream kinase responsible for ERK1/2 activation, suggesting an MEK-independent signalling mechanism. Pretreatment of PC3 cells with a PI3K inhibitor partially reduced both EGCG-induced ERK1/2 phosphorylation and cell proliferation inhibition. These results suggest that ERK1/2 activation via a MEK-independent and PI3-K-dependent signalling pathway is partially responsible for the antiproliferative EGCG effects in PC3 cells [20].

**2.6. Androgen Receptor (AR) and Prostate-Specific Antigen (PSA).** Experimental evidences suggest that androgens are



involved in PCa development and progression, being AR the essential mediator for androgen action. Detailed mechanisms of AR activation and modified function in PCa are reviewed in [37–39]. Briefly, the AR is a nuclear receptor activated through binding of its cognate ligands, such as testosterone and 5 $\alpha$ -dihydrotestosterone, and consequent dissociation by the heat shock proteins (normally bounded to it in the resting state). The activation process involves several coactivators recruitments. Activated AR up regulates the transcription of genes containing androgen response elements in their promoters as PSA gene that, specifically expressed in prostate, has been widely utilized for PCa screening, in the last 20 years [40]. A very recent report by means of a fluorescence resonance energy transfer (FRET) based assay provides evidence that EGCG is a direct antagonist of androgen action.

EGCG is capable to physically interact with the ligand-binding domain of AR by replacing a high-affinity-labeled ligand (IC<sub>50</sub>: 0.4  $\mu$ M) [41].

In different LNCaP sublines, EGCG suppresses cell proliferation, PSA expression, and AR transcriptional activity, at concentration comprised in the 10–20  $\mu$ M range [42, 43]. The effect on PSA expression might be related to reduction of AR activity, but it should also be considered, that EGCG, *in vitro*, can down regulate PSA by direct action on transcription and translation mechanisms [44, 45].

### 3. PCa Chemoprevention by Green Tea Polyphenols in Transgenic Mouse Model

Progress toward understanding the PCa biology has been slow due to the few animal research models of tumour onset and progression, available to study the spectrum of this uniquely human disease. Genetically engineered mice are being increasingly employed for delineating the molecular mechanisms of PCa development and the potential of new compounds as chemotherapeutic/chemopreventive drugs against it. Animal models present a rapid tumor growth comparing to the long latency of human PCa, which, on the other hand, makes the disease an ideal target for chemoprevention strategies. Mouse dorsolateral prostate lobe is functionally equivalent to human prostate peripheral zone, from where the majority of human cancer originate [46]. Preclinical studies with GTCs or with pure EGCG, administered at a human achievable doses, have been conducted in both transgenic animals and xenograft tumor models, in which murine and human cell lines, derived from primary tumor or metastasis, have been implanted subcutaneously [47, 48] or injected intraprostatically [49].

To the aim of studying human CaP, autochthonous murine models appear more suitable than orthotopic cell lines transplantation. In fact, transgenic mice exhibit sets of interactions between the different cellular, tissue and hormonal compartments appropriate to human prostate. Among the several lines of transgenic mice generated models, the transgenic adenocarcinoma of mouse prostate (TRAMP) has been well characterized and employed for a number of pre-clinical trials. Mice expressing the transgene display

progressive forms of prostatic disease that histologically resemble human PCa, from mild intraepithelial neoplasia (PIN) to poor differentiate adenocarcinoma phase (PD) and finally to metastatic spread [50]. In the TRAMP model, the SV40 early genes (T and t antigens, Tag) are under the control of the minimal rat Probasin promoter –426/+28—fragment [50], which renders the transgene expression androgen dependent, restricting it to the epithelial cells of the dorsolateral and ventral prostate lobes, thus abrogating p53 and Rb function [51] and inactivating protein phosphatase 2A (PP2A), specifically in this tissue [52].

In many experimental studies conducted by Gupta et al. [53] and by other authors [54–57], TRAMP mice aged from 8 to 32 weeks, received 0.1% oral infusion of a 95% GTCs enriched mixture. The animals, when compared to water-fed TRAMP mice, presented a significant delay in primary tumor incidence and almost complete metastases inhibition; prostate and genitourinary tract weight, a well-known tumor growth index, was decreased (64% and 72%, resp.), correlating with the reduced expression of Proliferating Cell Nuclear Antigen (PCNA).

The insulin-like growth factor pathway IGF/IGFBP-3 has been suggested to regulate PCa growth and development through its gradually increased activation during cancer progression.

After GTCs administration TRAMP mice showed a significant decrease in the IGF/IGFBP-3 ratio [54], accompanied by an inhibition of the downstream signaling cascade that involves both PI3K/AKT and MAPK pathways. Also, a parallel inhibition of vascular endothelial growth factor (VEGF), matrix metalloproteinases MMP-2, and MMP-9 expression were demonstrated [55]. Furthermore, the authors showed that metastasis-promoting Mts1 (S100A4) level, that, as a rule increases in cancer development, resulted in markedly decreased, E-cadherin level, that is progressively lost during cancer progression, where restored [56]. Also, the NF $\kappa$ B pathway activity, generally activated as a function of tumor grade, was reduced after 32 weeks of EGCG treatment, at a time when a shift in balance between Bax and Bcl2, favoring apoptosis, also occurred [57].

Under similar experimental conditions, we observed that, while 100% of TRAMP mice underwent PCa at 24 weeks of age, exhibiting tumor cell transendothelial passage in the absorbing lymphatic vessels, only 20% of the animals receiving 0.3% of GTCs (oral infusion), developed the neoplasm [19, 58–60]. In TRAMP mice presenting tumor growth arrest, a sequence of events were demonstrated, such as downregulation of H3 histone (a process usually affecting chromatin structure and gene expression), upregulation of growth arrest-Specific gene 1 (GAS1) and suppression of Mini-Chromosome Maintenance protein 7 (MCM7), a marker of DNA synthesis, essential for its replication, that is aberrantly expressed in various cancer types.

Interestingly, the level of the secretory protein clusterin (CLU) and mRNA, dramatically downregulated with the disease onset and development, resulted to accumulate progressively in the prostate after GTCs administration and remain undetectable in the 20% of animals that presented PCa, in spite of receiving GTCs solution. In correlation with

these observations, when tumor progression was inhibited, organelles committed to protein synthetic and secretory activities, as endoplasmic reticulum and Golgi apparatus, appeared significantly reduced in the prostate, suggesting possibly protein posttranslational processes alterations [60]. We suggest that CLU might participate in the chemopreventive action exerted by GTCs in TRAMP mice [19].

Harper et al. [61], after treating a cohort of TRAMP mice with almost pure 0.06% EGCG in drinking water, demonstrated that the compound can act only by slowing PCa progression. EGCG inhibited early (12-week-old mice), but not late (28-week-old mice) PCa stage. The treatment resulted in many various effects: AR, IGF1, and its receptor decreased level, apoptosis-reduced cell proliferation, and phospho-extracellular signal-regulated kinases 1/2, cyclooxygenase-2, and inducible nitric oxide synthase reduced activities. To a better interpretation of the data, it is worthwhile noting that green tea polyphenols bioavailability and transformation are key factors that can limit the compound activities *in vivo*; EGCG, as a single agent, may present a low bioavailability and/or slow its rapid metabolism, when stabilized by the naturally occurring mixture of green polyphenols. In addition, attention has to be paid when comparing experimental works on TRAMP mice colonies obtained through different background strains. In fact, the genetic background may have a profound effect on some aspects of tumor initiation/progression in this animal model.

The latest experimental work from Adhami et al. [62] confirmed the efficacy of 0.1% GTCs in drinking water to TRAMP mice, starting at defined stages of PCa onset and progression: 6 weeks (normal prostate), 12 weeks (PIN), 18 weeks (well-differentiated adenocarcinoma, WD), 28 weeks (poorly-differentiated adenocarcinoma, PD), finally cancer development. Tumor-free survival was indeed extended to 38 weeks in the 6 weeks group mice, but only to 24 weeks in the 18 weeks group, compared with 19 weeks in water-fed controls. Additionally, IGF and its downstream targets were significantly inhibited only when intervention was initiated at early stages.

The study design was conceived as a response to the outcomes of human clinical trials, based on alternative and complimentary green tea therapy, showing minimal effect on hormone-refractory PCa stage (CRPC) [63, 64], but lower incidence on high grade PIN stage [65, 66].

Extensive laboratory studies in multiple animal models consistently show the GTCs inhibitory activities against prostate carcinogenesis [14] and suggest the importance of identifying the most vulnerable PCa stages towards GTCs chemoprevention in humans.

#### 4. Possible PCa Chemoprevention by Green Tea Polyphenols in Humans

**4.1. PCa: Statistics and Importance for Chemoprevention Strategies.** PCa is the second most common cancer in American men with a 1 in 6 lifetime risk of developing it. The National Cancer Institute (NCI) estimated that approximately 217,730 new cases PCa would have been diagnosed

in 2010 and there would have been approximately 32,050 PCa deaths [67]. In Europe, PCa has become the most common non-skin cancer neoplasm among men, with an estimated 382,000 cases occurred in 2008 [68]. Incidence has increased rapidly over the past two decades, and rates are dramatically influenced by PSA testing, as well as by latent cancer detection in prostate surgery.

According to a recent report from the prostate, lung, colorectal, and ovarian cancer screening trial, six annual PCa screening programs (10 years followup results) led to increased number of diagnosed cancers, in the screening cohorts, but no cancer-specific survival advantage was seen for the group. Therefore, the real possibility to “overdiagnose” and “over-treat” cancers which are not life threatening, must be carefully taken in account [69]. A reasonable explanation for the low efficacy of invasive radical intervention, which are the gold standard in clinical practice after a diagnosis of confined PCa, relies on the fact that the disease is biologically heterogeneous and its natural history is almost unpredictable. Some cancers growth slowly, showing indolent course, while others are very aggressive, quickly progressing to advanced metastatic stage, that is almost an incurable disease [70]. Therefore, it is important and urgent to search for prevention strategy, effective when the disease is still at an early and potentially curable stage. PCa is an ideal candidate for chemoprevention, because it has a high prevalence, a long latency, and it is potentially lethal. Preventive strategies could carry a high economic benefit on the healthcare system reducing the costs associated with PCa diagnosis and therapy, moreover, it might have a deep positive impact on the patients quality of life, reducing the morbidity associated with radical surgery (incontinence and impotence) [71].

**4.2. Epidemiological Evidence of Green Tea Efficacy in PCa Chemoprevention.** PCa etiology is multifactorial, but the marked disparity in its incidence between “Eastern” and “Western” cultures suggests that dietary and lifestyle factors play an important role in the disease development and progression. This is strongly supported by migratory studies showing that Asian men, who relocate to the United States of America and adopt a western lifestyle, have a significantly higher PCa risk, when compared to their native Asian counterparts [72, 73].

During the last two decades, the relationship between tea consumption and cancer has been a subject of research interest for many investigators. Unfortunately at present, many epidemiological studies present conflicting results about the green tea role in cancer prevention. Recently, an exploratory meta-analysis of observational studies supported the hypothesis that green, but not black tea, may have a protective effect against PCa. A total of six epidemiological studies, including two case-control studies as well as four cohort studies, evaluated green tea role in reducing developing PCa risk [74–79]. A borderline statistical significant decrease in the disease development is presented with increasing green tea intake and, moreover, only case-control studies (odds ratio, OR = 0.43; 95% CI: 0.25–0.73), but not

prospective cohort studies (OR = 1.00; 95% CI: 0.66–1.53) reached a statistically significant result [80].

Negative and conflicting results in epidemiological data may be due to study design pitfalls and to many hardly controllable variables, like tea infusion composition and way of preparation (temperature), GTCs bioavailability, diet and the lifestyle of the people included in the study, and last but not least, genetic differences in the ability to metabolize GTCs.

**4.3. Pharmacokinetic Studies, Phase I Clinical Study, GTCs Metabolism and Tissue Distribution.** Polyphenon E is a GTCs-enriched and defined product, virtually caffeine free (<0.5%) produced by Mitsui Norin Co. Ltd, a new drug investigated by the Food and Drug Administration. It contains 80% to 98% total catechins by weight, with EGCG main component, accounting for 50% to 75% of the material.

Chow et al. [81, 82] performed several pharmacokinetic phase I studies (in healthy volunteers) utilizing Polyphenon E (capsules) to determine the systemic tea polyphenols availability after a single or multiple dose and various dosing condition (200, 400, 600, 800 mg of EGCG). In the single-dose study, Polyphenon E was compared to pure EGCG for differences in pharmacokinetic parameters [81]; after its administration, at the four dose levels above mentioned, the average EGCG peak plasma concentration ( $C_{max}$ ) was  $72.7 \pm 66.4$ ,  $125.3 \pm 50.4$ ,  $165.7 \pm 126.9$ , and  $377.6 \pm 149.8$  ng/mL. Similar results were obtained after pure EGCG administration, at the same concentration, suggesting that GTCs and Polyphenon E do not have any effect on the EGCG pharmacokinetics. It should be noted that tea catechins bioavailability is quite low in humans, resulting in plasma concentrations 5 to 50 times less than concentrations shown to exert biological activities *in vitro* [14].

In the multiple dose study, the same authors evaluated effects and safety following chronic Polyphenon E administration [82], concluding GTCs do not accumulate in the body and are safe for human health.

A third pharmacokinetic study from the same authors [83], evaluated the role of fasted or fed state on Polyphenon E bioavailability (400, 800, or 1200 mg of EGCG), concluding that  $C_{max}$  of free catechins (EGCG, EGC, EC) can be increased of about 3-folds, when administered in a fasted state. The first study demonstrating that GTCs can be detected in the prostate tissue after green tea consumption has shown that after a short period (5 days) of green tea continuous intake (1.42 L of brewed tea divided in 5 daily doses), the prostate GTCs concentration were 0.1, 0.043, 0.040, 0.0021 (nmol/g tissue) for EGC, EC, EGCG, and ECG, respectively [84].

**4.4. Proof of Principles and Phase II Studies of Green Tea Efficacy for PCa Chemoprevention and Treatment.** There have been 5 intervention studies evaluating the GTCs effect on PCa treatment or prevention [63–65, 85, 86]. Three were single-arm open-label phase II trials, performed in patients diagnosed with a castration resistant PCa (CRPC) [63, 64, 86] and one was a pilot proof of concept study, performed

to evaluate GTCs ability to reduce cancer incidence in a well-defined cohort of patients, bearing premalignant lesions (HGPIN) [65].

The most recent study, a randomized, double-blind, placebo-controlled phase II trial evaluated the effect of short-term Polyphenon E administration in men with PCa scheduled to undergo radical prostatectomy [85].

The primary endpoint of the trial reported in [64] was to determine the capacity of nonstandardized green tea powder to produce a PSA base level decline in 42 men with clinical CRPC evidence. Only 1 patient, among 42, manifested a transitory 50% decreased PSA level from baseline, during the 6 months followup. The negative result of this study needs to be critically considered; it should be taken into account that the enrolled population comprised men with an advanced cancer stage, that acquired resistance to standard hormone deprivation therapy, and that a nonstandardized GTCs preparation was employed.

Choan et al. [63] published the results of a study aimed to evaluate the effect of a standardized green tea extract (250 mg GTCs/day) on PSA level or measurable marks of the disease progression, after a minimum of 2 months therapy. Only 15, out of the enrolled 19 patients, completed at least 2 months therapy and all of them exhibited a progressive disease in the first 4 months. In addition, a very small population with an advanced cancer stage, unresponsive to previously administered hormonal therapy, was considered.

On the basis of encouraging results obtained by us and others in the PCa chemoprevention with animal models [19, 53, 62], our research group performed a proof of concept trial in a well-selected patients cohort at high risk to develop PCa [65]. We enrolled 60 patients with HGPIN diagnosis, that were randomly assigned to receive, according to a double-blind procedure, a standardized GTCs formulation (600 mg/day), or identical placebo capsules for 1 year. GTCs composition, virtually caffeine free, was total catechins 75.7%; EGC, 5.5%; EC, 12.2%; EGCG, 51.9%; ECG. All the patients, during the study, were subjected to regular prostate biopsy (6 and 12 months after the study beginning) to assess differences in PCa values between the two arms. Total serum PSA concentration was measured at 3, 6, 9, and 12 months, to check whether GTCs administration would reduce the value. Because concomitant benign prostate hypertrophy (BPH) was very common among the patients enrolled, we also evaluated the International prostate symptom score (IPSS) and quality of life (QoL) after 3 months of GTCs administration in a subcohort of patients with low urinary tract symptoms (LUTSs), as a further secondary end point. We found that 9 out of 30 patients receiving placebo developed PCa, while only 1 out of 30 patients receiving GTCs was found PCa positive after prostate biopsy. We did not find any significant difference in PSA values between the 2 arms, but we found a significant IPSS and QoL scores improvement.

Our results point the attention on the fact that GTCs should be considered interesting and promising natural compounds for PCa chemoprevention, while they are almost ineffective in case of full-developed metastatic neoplasia (CRPC). We also found that PCa incidence remained low



in patients belonging to the GTCs-arm, even two years after therapy suspension [66]. This result may suggest that chemoprevention activity and clinical benefits achieved with GTCs are stable over time. We hope that our data will be confirmed by a large phase II study, now ongoing, sponsored by NCI. This trial is aimed to enroll about 300 HGPIN patients to be given 400 mg/day of Polyphenon E for PCa prevention. (Study of Polyphenon E in men with high-grade prostatic intraepithelial neoplasia, Protocol IDs: MCC-15008, R01 CA12060-01A1, NCT00596011).

McLarty et al. [86] published the results of a study aimed to evaluate the effect of short term (median treatment period 34.5 days) GTCs administration in a population of PCa diagnosed patients, scheduled for radical prostatectomy. The supplementation, performed with an high, well-tolerated, Polyphenon E dose (1.3 g GTCs/day) produced a significant decrease of biomarkers relevant for PCa development, like HGF (hepatocyte growth factor), VEGF (vascular endothelial growth factor), IGF-1 (insulin growth factor-1) and IGFBP-3 (insulin growth factor binding protein-3). These results support a potential GTCs role in PCa chemoprevention and treatment in early confined stage.

Disappointingly, the positive Polyphenon E influence resulted not statistically significant according to the results of a randomized, double-blind, placebo-controlled trial, having a study design strictly close to that of McLarty [85]. The reason(s) for the discrepancy between the two studies remains to be elucidated, but it should be considered that the lack of a control group in the single arm study made it easier to gain statistical significance. On the other hand, it may indicate that future studies would need a larger population to show a statistically significant difference in systemic biomarkers and Polyphenon E, or generally standardized GTCs formulation, should be preferentially tested in longer-term intervention studies and in a precancerous model, where its effects have a best chance to be demonstrated.

## 5. Molecular Mechanisms for Green Tea Polyphenols Anticancer Activity in PCa: An Emerging Role for Epigenetics

**5.1. Modulation of Epigenetic Mechanisms.** In the last five years, GTCs have been shown to be able to modulate all the principal epigenetic mechanisms: DNA methylation, regulation of chromatin structure (through histone posttranslational modifications), and alterations of noncoding miRNAs. Epigenetics is defined as changes in gene expression that do not involve changes in the DNA sequence. Importantly, these changes are both reversible and heritable through division of somatic cells, making epigenetic regulation of gene expression a dynamic process that plays a crucial role in a vast number of biological processes including development, cell differentiation, stem cell maintenance, and tissue homeostasis [87]. Deregulation of epigenetic mechanisms is found in numerous diseases and in virtually every kind of cancer. This observation, together with the dynamicity, and therefore the potential reversibility, of epigenetic mechanisms sparked the interest in developing epigenetic drugs capable to modulate

DNA methylation and chromatin structure in cancer cells. In the last few decades, several epigenetic drugs have been developed and are now either tested in clinical trials or already established in the clinical care, even if the drugs, besides modulating epigenetic mechanisms, appear to exert a certain degree of cytotoxicity.

The observation that GTCs, together with other dietary polyphenols, can regulate specific epigenetic features of premalignant and malignant cells opens new possibilities for epigenetic therapy

**5.2. DNA Methylation.** DNA methylation is an important epigenetic determinant in gene expression. As a matter of fact, it participates in the maintenance of DNA integrity and stability, in chromosomal modification, and development of mutations. Generally, DNA hypermethylation is associated with genes inactivation and global genomic hypomethylation is associated to chromosomal instability induction. The DNA methylation occurring on the carbon-5 position of cytosine residues within a CpG dinucleotide sequence, represents the most studied epigenetic marker. In normal tissues, the process shows a bivalent function: CpG sites clustered in regulatory regions, within promoters and enhancers (CpG islands), are usually unmethylated, allowing for genes to be expressed; while sparse CpG sites (throughout the genome) are usually methylated, contributing to genome stability [88]. DNA methylation is crucially dysregulated in the vast majority of cancers, where CpG islands become hypermethylated, causing silencing of many genes, involved in cell cycle regulation, tumor suppression, DNA repair enzymes, receptors activity, and apoptosis, while the sparse CpG sites are usually subject to hypomethylation, favoring genomic instability, a common feature of cancer etiology [89].

Epigenetic analysis offers a potential noninvasive blood marker, complementary to PSA, for a preliminary PCa diagnosis. Since inhibition of DNA methyltransferases (DNMTs) may prevent hypermethylation and silencing of tumor suppressor key genes, it is reasonable to suppose that enzyme inhibition, along with histone deacetylase activation, may contribute to cancer treatment and or carcinogenesis prevention.

In human prostate cancer PC3 cells, as well as in other cell lines, Fang et al. [90] demonstrated that EGCG decreases total DNMTs activity, leading to reactivation of several genes silenced by methylation, such as retinoic acid receptor beta (RAR $\beta$ ), which expression results increased by lowering the methylation levels of its promoter. This is the first demonstration of such an activity of a commonly consumed dietary constituent.

Recently, it has been reported that exposure of human prostate cancer LnCaP cells to GTCs causes a time- and concentration-dependent reactivation of the expression of the glutathione-S-transferase p1 gene (GSTP1) [91]. The gene, mostly studied for methylation in PCa, results hypermethylated in almost 90% of the tumors [92]; in fact, it is recognized as a molecular PCa hallmark. Cells treatment with GTCs results in GSTP1 promoter demethylation, associated with a significant reduction of DNMT1 activity. Interestingly, the treatment does not cause global sparse CpG sites

hypomethylation, contributing to maintain genome stability. The authors present GTCs as excellent candidates for epigenetic chemoprevention against PCa, even more favorable than the most commonly DNMT inhibitors employed, such as 5-aza-2'-deoxycytidine.

There is growing evidence that the epigenetic mechanisms that impact DNA methylation and histone status, also contribute to genomic instability. Instead, GTCs administration lacks of toxicity, do not reactivate the prometastatic gene S100P, as a reverse response of 5-aza-2'-deoxycytidine administration, and is able to alter chromatin modeling by histone acetylation (the second global epigenetic mechanism of gene regulation, see below) [91].

Almost at the same time, Morey Kinney et al. [93] showed that GTCs administration to TRAMP mice (spontaneously developing prostate adenocarcinoma) had very little effect on DNA methylation. They have found that 5-methyl-deoxycytosine level, together with methylation levels of B1 repetitive elements and the Mage-a8 gene, that are correlated to the phenomenon, remained unchanged in wild-type and TRAMP mice prostate. Also, they performed a genome-wide DNA methylation profiling of the HpaII tiny fragment enrichment by ligation-mediated PCR (HELP) [94] and found no significant hypomethylation. Their study, however, arises some doubts on the interpretation of the data: most of the analysis is directed towards sparse CpG site, rather than CpG islands. Indeed, the lack of global hypomethylation in GTCs fed mice agree with the above results obtained in LnCaP cells [91]. Second, but most important, in their study, GTCs treatment did not cause chemopreventive effects on PCa development, in contrast with previous reports by other authors [19, 53]. The different composition of the GTCs preparation used by Morey Kinney et al. [93] might account for this discrepancy, since it contained only about half of the EGCG concentration (35%) relative to the other polyphenols, when compared to the preparations used in other studies (51.9% in [19] and 62% in [53]).

Among the still growing number of experiments aimed to clarify the GTCs power as epigenetic drugs against PCa, there is plenty of evidence that the GTCs can interfere with DNA methylation pathways *in vitro*, leading, also, to the reactivation of the tumor suppressor p16, known to be involved in the cell cycle regulation [95].

Three different molecular mechanisms, at least, have been proposed to explain GTCs effects on the DNA methylation process (Figure 2). In human models, GTCs are demonstrated to be readily methylated, by catechol-O-methyltransferase (COMT) which utilizes S-Adenosyl-L-methionine (SAM) as a methyl donor, producing equimolar concentration of S-adenosyl-L-homocysteine (SAH). It should be noted that SAM is an indispensable methyl donor for DNMTs and SAH is a potent inhibitor of DNMTs activity. Since GTCs administration causes both decreased SAM and increased SAH cellular concentrations, the hypothesis is proposed that GTCs may act by perturbing the regulatory physiological homeostasis of the two molecules [96].

This mechanistic explanation is supported by Lee et al. [97]. The authors, utilizing multiple modelling tools, have shown that the inhibitory effect exerted by several

polyphenols on human DNMT1 is increased after *in vitro* COMT addition. In these experiments, EGCG resulted the much more potent DNMT1 inhibitor, able to act also independently of the COMT presence. In addition to the indirect action on DNA methylation demonstrated by [96], EGCG is able to inhibit DNMT1 by directly binding to the enzyme [90], with the gallic acid moiety of the molecule playing a crucial role in the interaction (Figure 2).

In human cell lines, EGCG is shown to act as an antifolate compound and disturb the folic acid metabolism by inhibiting dihydrofolate reductase. Since folic acid physiologically modulates DNA methylation, Navarro-Peran et al. [95] hypothesize that many molecular effects imparted by EGCG administration might be explained by this kind of interference with DNA methylation (Figure 2).

**5.3. Chromatin Structure.** Changes in the chromatin condensation, a crucial mechanism to control gene expression in eukaryotic cells [98], is regulated by at least eight posttranslational modifications of histones, such as acetylation, methylation, phosphorylation, poly-ADP-ribosylation, ubiquitination, glycosylation, and so forth, predominantly occurring on lysine residues [99]. Combinations of the different modifications constitute the histone code that defines the actual or potential transcriptional state [100] through proteins-DNA interaction, which in turns regulates gene expression.

Among the most studied processes, histone acetylation/deacetylation is finely regulated by balancing the activity of acetyl-transferases (HATs) and histone deacetylases (HDACs) enzyme families; inhibition of the latter rapidly results in histone hyperacetylation. Acetyl groups added to lysine residues by HATs, mainly on H3 and H4 histones, neutralizes aminoacid positive charge and consequently loosens DNA binding to histone complexes, ultimately resulting in chromatin relaxation and gene activation [99].

Being HDACs overexpression implicated in protecting cancer cells from genotoxic insults, many HDAC inhibitors synthesized in order to favor tumor suppressor genes re-expression and silence DNA repair pathways, are gaining a momentum as a novel cancer therapy. Raiendran et al. [101], however, indicate how dietary phytochemicals, by affecting the epigenome, and also can trigger DNA damage and repair mechanisms.

With regard to PCa, Pandey et al. [91] have observed a time-dependent inhibition of the total HDACs activity, after GTCs administration to LnCaP cells, correlating with mRNA and HDAC1-2-3 classes decrease.

Recently, GTCs administration was shown to inhibit HDAC1 activity and protein expression in PCa cell lines as LnCaP (harboring wild-type p53) and p53-null PC3 (lacking p53). When administered together a proteasome inhibitor, GTCs effect resulted prevented, indicating that HDAC1 is proteasoma degraded. HDAC1 inhibition was followed by up regulation of the cell cycle inhibitors WAF1/p21, ultimately inducing G<sub>0</sub>/G<sub>1</sub> phase cell cycle arrest and apoptosis in both cell lines, irrespective of their p53 status [102].

A sustained DNA damage response, coupled with insufficient repair mechanisms, may be a pivotal mechanism

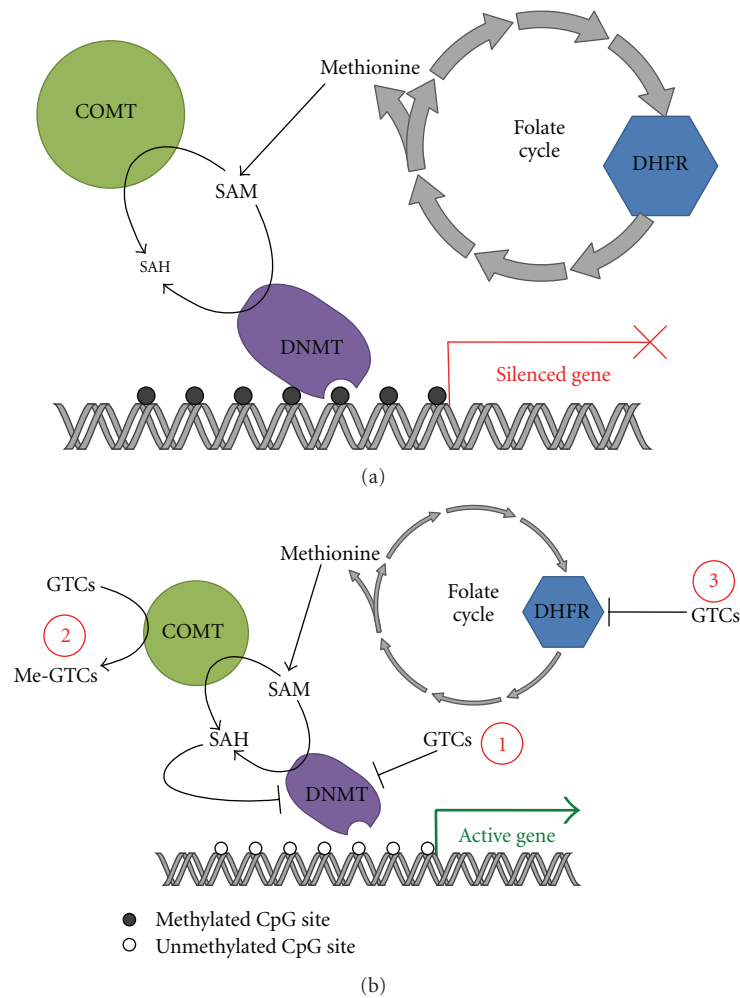


FIGURE 2: Mechanisms of inhibition of DNMTs by GTCs. (a) In a cancer environment, specific genes are silenced by hypermethylation of their promoters. DNA hypermethylation is catalysed by DNMTs, which use as a substrate SAM and release SAH as a by-product. (b) Within this context GTCs are able to inhibit DNMTs through three distinct mechanisms. (1) Direct inhibition. (2) GTCs are methylated by COMT, resulting in a depletion of SAM and accumulation of the DNMT inhibitor SAH. (3) Direct inhibition of DHFR, resulting in disruption of the folate cycle that influences negatively the levels of SAM. Inhibition of DNMTs ultimately results in DNA hypomethylation and re-expression of previously repressed genes.

for apoptosis induction in cancer cells exposed to dietary phytochemicals. The above findings provide new insight into the GTCs mechanisms, suggesting a novel approach to prevention and/or therapy.

**5.4. microRNAs.** microRNAs (miRNAs) comprise a novel class of endogenous, small, noncoding RNAs that control gene expression by acting on target mRNAs for degradation and/or translational repression. Transcription of miRNA genes and protein-coding genes shares common regulatory mechanisms: miRNA genes can be embedded in the introns of protein coding genes or can derive from their own transcript units in intergenic regions of the genome. When miRNA genes are located within introns of protein-coding genes, primary miRNA biogenesis is controlled by the same transcriptional mechanisms as the parent gene. In contrast, an independent miRNA gene will have its own

transcriptional controls. Interestingly, multiple miRNAs can be produced within a single transcript, each of which can act independently [103, 104]. The long primary nuclear miR transcript ("pri-miRNA") undergoes maturation by the RNase-III Drosha/Dgcr8 enzyme complex, generating a precursor miRNA ("pre-miRNA") that is exported from the nucleus by exportin-5 [105]. A second cleavage takes place in the cytoplasm, involving the action of a complex containing another RNAase III enzyme named Dicer [106], to generate a single-strand mature miRNA. Mature miRNA, is usually incorporated in RNA-Induced Silencing Complex, a complex of proteins that is responsible for silencing the target mRNA [107]. Translational inhibition or degradation of targeted mRNA transcripts is due to imperfect or perfect base pairing between positions 2 to 8 from the 5' miRNA (also known as the seed sequence), with the 3' Un-Translated Region of their target mRNAs [104].

miRNAs are emerging as important regulators of gene expression and we expect them to be key players in intracellular signalling, thus enabling to close gaps of knowledge in molecular pathways. Pathogenic roles of miRNAs were initially described in cancer and miRNAs have now become a hot topic in medical research [108].

**5.5. microRNA in PCa.** To our knowledge, by employing miRNA microarray analysis, 7 miRNAs (miR-145, miR-141, miR-125, miR-1, miR-133, miR-106b, and miR-16) have been found downregulated in PCa [109–111]. In addition, the chromosomal region containing the miR-15/16 cluster and miR-101 is often lost during PCa progression. Table 1 summarizes recent publications on miRNAs in PCa.

Bonci et al. [112] demonstrated that reduced levels of miR-15/16 are associated with PCa growth and due to an increase in the protein levels of their target genes Bcl2, cyclin D1, and WNT3A. Conversely, overexpression of miR-15/16 suppresses tumor growth and induces its regression. Expression of miR-101 inversely correlates with upregulation of its target, enhancer of zeste homolog 2 (Ezh2), which is highly expressed in CRPC [113].

miR-331-3p and miR-449a are downregulated in PCa, contributing to cancer growth by overexpressing their targets ErbB2 [114] and HDAC1, respectively [115]. Poliseno et al. [116] demonstrated that miR-22 and the miR-106b~25 cluster are overexpressed in PCa, and potentiate cellular transformation both *in vitro* and *in vivo*. Intronic miR-106b~25 cluster cooperates with its host gene *MCM7* in cellular transformation both *in vitro* and *in vivo*.

Other tumor suppressor miRNAs in PCa are miR-34 especially in p53-deficient PCa cells [117] and miR-330, which suppresses E2F-1 and E2F-1-mediated AKT phosphorylation [118]. In PCa cells, miR-21 is induced by stimulation of androgen receptors and mediates hormone-dependent and -independent cell growth [119]; in contrast, miR-221 increases in androgen-independent tumors [120]. MiR-146a, which is reduced in androgen-independent PCa cells, inhibits proliferation and cell invasion by targeting of Rho-activated protein Kinase 1 [121].

miRNAs that have been detected in human serum and plasma specimens, and circulating miRNA profiles, have now been associated with cancer, as an emerging class of diagnostic and prognostic biomarkers. Specifically in PCa, the expression of miR-141 has been found to be elevated in the plasma of PCa patients [122], where levels of miR-141 in PCa were able to predict tumor progression, when compared with other validated prostate biomarkers [123]. Finally, miR-141, miR-298, and miR-375 were also found to be upregulated at differential levels in the serum of men with CRPC [124].

**5.6. microRNAs in PCa and Polyphenols.** Recently, studies support a growing interest in the chemopreventive role of dietary agents such as polyphenols, as modulators of miRNA profiles in cancer progression and prevention [125]; (Table 2).

TABLE 1: miRNAs involved in PCa.

miRNAs	Target genes	References
miR-15/16	Bcl2, CCD1, WNT3a	[112]
miR-101	Ezh2	[113]
miR-331-3p	Erb2	[114]
miR-449a	HDAC1	[115]
miR-146a	ROCK1	[121]
miR-106b/25	PTEN	[116]
miR-330	E2F1	[118]

TABLE 2: miRNAs modulated by polyphenols in PCa.

miRNAs	Polyphenols	Regulation	References
miR-21	EGCG	down	[41]
miR-330	EGCG	up	[41]
miR-1296	Genistein	up	[129]
miR-17/92	Resveratrol	down	[130]
miR-106a/b	Resveratrol	down	[130]

Although the chemopreventive effects of EGCG, as well as other polyphenols, have been largely demonstrated in PCa by us [19, 59, 126] and other groups (reviewed in [127]), only few reports discussed the regulatory effect of miRNA expression by dietary components in PCa. At the moment, the first and only study that clearly correlated miRNA-regulation with catechins treatment in PCa is performed by Siddiqui et al. [41]. The authors studying the role of androgen receptor (AR) in both early and advanced stage of PCa etiology, showed that the androgen-regulated miRNA-21 and the tumor suppressor miRNA-330, commonly regarded to play a role in PCa, are, respectively, down- and upregulated in a xenograft mice model for PCa, following EGCG treatment. These findings strengthen EGCG as an androgen receptor signalling antagonist, which can block AR gene expression and cell growth in the human PCa cells. EGCG is suggested as a chemotherapeutic agent against CRPC.

Basing on experiments performed in pancreatic cells with the phytochemical substance curcumin by Bao et al. [128], it appears worthwhile investigating the EGCG effect on miR-200 and miR-21 expression in PCa cells. Interestingly, the increase of the two above miRNAs is correlated to the induction of the critical tumor suppressor gene PTEN, frequently defective in PCa.

A study examined the effect of the phytochemical compound genistein on minichromosome maintenance (MCM) genes, commonly dysregulated in cancer, showing that MCM2 expression genes is higher in PCa samples, whereas miR-1296 was significantly downregulated [129]. Genistein induced miR-1296 expression and subsequently downregulated the MCM2 expression, along with cell cycle arrest in S-phase. This study is worth of our attention, since we demonstrated a dramatic MCM7 genes suppression by GTCs in the prostate of TRAMP mice [59]; analyses of therapeutic effect of catechins on miR-1296 and on miR-106b~25 cluster, request further investigations.



TABLE 3: Principal progressive phases in human PCa development.

Definition	Acronyms	STAGE of PCa development	STAGE where CTCs could be possibly used
Proliferative Inflammatory Atrophy	PIA	Precursor of cancer initiation	YES
Prostatic Intraepithelial Neoplasia	MILD PIN HIGH PIN	PreMalignancy Malignancy	YES YES
Adenocarcinoma	WD PD	Well-differentiated Cancer POOR differentiated Cancer	NO NO
Fully Developed Metastatic Neoplasia	CRPC	Hormone refractory castration resistant Terminal cancer	NO

Finally, utilizing miRNA microarrays, Dhar et al. [130] found that miR-17-92 and miR-106ab clusters, with well-recognized oncogenic properties, were significantly down-regulated after treatment with the phytochemical compound resveratrol.

To corroborate the hypothesis that EGCG may act as a negative tumorigenesis process, downregulating oncogenes miRNAs and/or upregulating tumor suppressor genes miRNAs, more definitive informations are needed.

## 6. Conclusion and Future Perspective

Despite the routinary employment of intermediate-risk prognosticators such as PSA, Gleason score, and T-category, PCa remains a complicate malignancy, exhibiting high heterogeneity features both when present in latent, clinically indolent, and in progressively more aggressive stages, leading to the “hormone refractory state” (CRPC). This is the most devastating PCa form, representing the terminal stage of transition from the androgen dependence stage, against which, at the moment, no curative therapy exists.

Although radical prostatectomy is curative in the majority of patients with clinically localized PCa, up to 40% of them fail to respond to local therapy and develop PSA recurrence, as a sign of metastatic growth; ultimately, many of these patients die from their disease. While latent PCa appears similarly frequent in men with different ethnic background, residing in culturally diverse geographic locations, experimental evidences show that the quality of diet and lifestyle factors may carry the mechanisms triggering the transition to the “hormone-refractory state” and, at the same time, may be the target for the action of anticancer prevention.

These considerations support the increasing use of complementary and/or alternative therapies such as developing a diet-based combinatorial approach. Chemopreventive action of a naturally occurring, nontoxic agent, such as green tea polyphenols, could be useful in the PCa management. Just a postponement of the “hormone-refractory state” or the maintenance of the androgen dependence would produce chronic instead of terminal PCa. On the other hand, there is growing consensus that a large subset of patients do not require aggressive treatment. Recent preclinical evidences showing that GTCs can inhibit cell cycle, induce apoptosis, and modulate several signalling pathways, strengthened by

experiments with animal models, have demonstrated the possible utilization of these compounds in selected human PCa stages for preventing its development (Table 3).

To date, although additional investigations are needed, a real cancer preventive activity by tea polyphenols has not been consistently observed in the few studies with humans versus the several studies with animal models so far achieved. Table 4 suggests two main differences between humans and animals studies: (1) relatively low quantities of GTCs intake by patients, as compared to TRAMP mice (it is conceivable that GTCs availability in the prostate is different in mouse than in humans; on the other hand pharmacokinetics studies demonstrate EGCG limited systemic concentrations); (2) various confounding factors (as genetic differences, diet, lifestyle and, etc.) present during the patients treatment, unlike the controlled conditions to which animals are subjected to optimize cancer prevention effect.

In the future, efforts will be needed to monitor GTCs administration to humans starting from the standardization of optimal GTCs compositions and doses, and continuing by enhancing GTCs bioavailability through nanocapsules or liposome deliver (nanochemoprevention). Simultaneous check of plasmatic and bioptic EGCG levels will be indispensable; it remains to be determined whether EGCG undergo autooxidation in the prostate gland, as it occurs in cell culture.

Certainly novel biomarkers focused not only on PCa epithelial cells but also on cell interaction with the extracellular microenvironment, that will improve the ability to detect PCa, predict lethality, and monitor response to therapies (many markers validated in experimental studies are not yet introduced as routine diagnostics).

Substantial information on tea polyphenols chemopreventive activity will emerge from long-term and rigorous clinical trials on well-designed cohorts of patients. Although it may appear simplistic that clinical PCa heterogeneity is attributable to underlying molecular heterogeneity, clinical trials still have to consider the need of cohorts with definite cancer risk, classified by their gene expression signature, that represents a clinically relevant genetic biomarker, independent of the current diagnostic variables. PCa develops via a limited number of alternatively preferred genetic pathways, providing genetic subtype (molecular classification) that may constitute a framework for investigating PCa and possibly explain the clinical heterogeneity of the disease with regard to both tumor progression and therapeutic response.

TABLE 4: Studies on the effect of GTCs administration to TRAMP mice and humans with PCa.

Experimental groups	GTCs dose g/100 mL [oral administration]	Formulation (%)	PCa phases age-dependent (weeks)	PCa inhibition (%)	References
TRAMP mice	0.06	<b>GTCs<sup>a</sup></b> EGCG (93)	Start 5; end 12	83	[61]
	0.1	<b>GTCs<sup>b</sup>:</b> EGCG (62) ECG (24) EGC (5) EC (6)	Start 8; end 32	42	[53]
	0.1	<b>GTCs<sup>b</sup>:</b> EGCG (62) ECG (24) EGC (5) EC (6)	Start 6; end 38	~50	[62]
	0.1	<b>GTCs<sup>b</sup>:</b> EGCG (62) ECG (24) EGC (5) EC (6)	Start 18; end 24	~20	
	0.3	<b>GTCs<sup>c</sup>:</b> EGC (5.5), EC (12.2) EGCG (51.9) ECG (6.1)	Start 8; end 24	80	[19]
	GTCs dose mg/day [oral administration]	GTCs providers	PCa phases	Percentage (%) of PCa inhibition	References
HUMANS	500	Sabinsa Corporation	CRPC	No effect	[63]
	6000	Unilever	CRPC	No effect	[64]
	1300	Polyphenon E Matsui Norin,	CRPC	Mild effect	[86]
	600	Polyphenon E Matsui Norin	HGPIN	33	[65]
	800	Polyphenon E Matsui Norin	CRPC	No effect	[85]

GTCs<sup>a</sup>: Roche, GTCs<sup>b</sup>: natural resources and products, GTCs<sup>c</sup>: isolated by the investigators; CRPC (castration resistant prostate cancer), HGPIN (high grade prostatic intraepithelial neoplasia).

Conclusively, to avoid difficulties in the interpretation of the data, the future tea polyphenols protocols should take into account the following: (1) standardization in the preparation of the substances employed, choice of their doses and concentrations, systematically tested in plasma as well in biopsies; (2) more accurate stratification of a large number of the patients utilized for the analysis and selection of an appropriate treatment duration.

## Abbreviations

AR: Androgen receptor  
 PCa: Prostate cancer  
 EC: Epicatechin  
 ECG: (-)-Epicatechin-3-gallate  
 EC: (-)-Epicatechin  
 EGC: (-)-Epigallocatechin  
 EGCG: (-)-Epigallocatechin-3-gallate  
 GTCs: Green tea catechins  
 miRNA: MicroRNAs

ROS: Reactive oxygen species

PSA: Prostatic specific antigen

TRAMP: Transgenic adenocarcinoma of mouse prostate.

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